Diversity of Zygomycetes associated with the rhizosphere of woody plants from Ile-Alatau national park (Kazakhstan)

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Abstract

Total 9 species of Zygomycetes belonging to six different genera, were identified from the rhizosphere of various woody plants in Ile-Alatau state national Natural Park. *Mucor mucedo* was observed as the most prevalent Zygomycete species. This species was isolated from the rhizosphere of apple, apricot, aspen, birch, oak, pine, poplar, spruce, rowan and willow. In addition to *Mucor mucedo* three additional species of the genus Mucor were noted: *M. fuscus* isolated from the rowan rhizosphere, *M. silvaticus* found in the apple rhizosphere, *M. strictus* was detected in the birch rhizosphere. *Absidia spinosa*, isolated from the rhizosphere of apple, apricot, hawthorn, inn, poplar, rowan, spruce and willow, was the next prevalent fungus in the studied rhizosphere samples. Only one species of *Cunninghamella, C. echinulata*, from hawthorn rhizosphere was detected. *Actinomucor elegans* was isolated also from the hawthorn rhizosphere. *Piptocephalis arrhiza* isolated from the apricot rhizosphere, grown together with species of the genus *Mucor*, as their parasite. *Rhizopus stolonifer* was isolated from the spruce rhizosphere.

Eight species of Zygomycetes are found in the rhizosphere of mixed forest trees, three species are found in the soil of spruce forests. Two species are common to mixed and spruce forests. The greatest number of soil fungi species is typical for mixed and small-leaved forest at the altitude range 1400–2000 m above sea level, much less of them in higher ranges. *Absidia spinosa* and *Mucor mucedo* are dominant for these altitudes. The number of soil Zygomycetes varies in the rhizosphere of different forest species. The greatest number of soil fungal species (3 species) is observed in the rhizosphere of apple, apricot, birch, hawthorn, rowan and spruce.

Key words – columella – coniferous forest – fungal colony – merosporangium – mixed forest – occurrence frequency – small-leaved forest – sporangiospore – sporangium – zygospore

Introduction

Ile-Alatau state national Natural Park was established by the decree of the Government of the Republic of Kazakhstan on February 22, 1996 to preserve the unique landscapes, flora and fauna. The Park's area is 198,669 hectares. National park is located on the Northern macroslope of Zailisky Alatau, the most Northern range of the Tien Shan. Range is almost 380 km long, 30-40 km wide. The highest peak Talgar reaches 5017 m above sea level. The climate is diverse and differentiated by high-altitude climatic zones. Vegetation is characterized with specific spectrum of altitudinal belts. There is no continuous belt of coniferous forests and alpine tundra belt. A distinctive feature of Zailisky Alatau ridge is the wide spread of all subtypes of mountain steppes.
In mycological respect, Zailisky Alatau ridge is sufficiently studied. A variety of ecological
conditions of the ridge, as well as the peculiar composition of host plants, could not affect the
number and ratio of different groups of fungi, which are taken into account here more than 2000
species (Schwarzman 1962). However, the soil mycobiota of has been poorly studied. The list of
soil fungi species in the mixed forests of Zailisky Alatau consisted of only 4 species of
micromycetes, 20 species were found in coniferous forests (Schwarzman 1962). To our research
Zygomycetes is not marked in the soils of the national park.

The purpose of this study was to add species of Zygomycetes to the list of soil fungi that have
been detected in the forests of Zailisky Alatau, to provide short descriptions, remarks on the
species, descriptions of the fungal colonies, and microphotographs.

Materials & Methods

Sampling sites
Soil samples of the forests of the Ile-Alatau national Park (Fig. 1) were collected during the
growing season of 2018 (April, July, and August). At the sampling sites the rhizosphere of the trees
of dark coniferous, small-leaved and mixed forests on mountain-forest soils in Zailisky Alatau were
sampled. Dark coniferous forests are composed of the main forest-forming species of the Shrenk
spruce (*Picea schrenkiana* Fisch. & C. A. Mey.). Underwood consists of rowan, aspen, various
species of rose hips, honeysuckle, juniper, and willow. Lower zone in Zailiyksy Alatau and valleys
of the rivers are occupied by the mixed deciduous-fir woods. In addition to spruce, the dominant
species were apple, apricot, willow, birch, aspen.

![Fig. 1 – Map showing soil collection sites in central part of Ile-Alatau National Park](image)

Soil sampling
Samples were collected randomly in the soil horizon of 5-20 cm, after removal of the top
litter layer, in the rhizosphere of various woody plants. The geographic location of each sample
collection site was recorded using digital GPS (Germin). In total, 44 samples were collected in three replicates, from which 35 species of soil fungi were isolated, among them 9 species of Zygomycetes (from 25 samples) (Table 1).

**Table 1** Data on soil samples from which Zygomycetes were isolated

<table>
<thead>
<tr>
<th>Samples</th>
<th>Altitude above sea level</th>
<th>Latitude, N</th>
<th>Longitude, E</th>
<th>Rhizosphere of</th>
</tr>
</thead>
<tbody>
<tr>
<td>256</td>
<td>1924</td>
<td>43°06’24.1&quot;</td>
<td>76°57’12.8&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>257</td>
<td>1927</td>
<td>43°06’23.8&quot;</td>
<td>76°56’46.4&quot;</td>
<td>poplar</td>
</tr>
<tr>
<td>258</td>
<td>1930</td>
<td>43°06’28.7&quot;</td>
<td>76°56’21.2&quot;</td>
<td>poplar</td>
</tr>
<tr>
<td>262</td>
<td>1926</td>
<td>43°06’27.1&quot;</td>
<td>76°56’15.9&quot;</td>
<td>apricot</td>
</tr>
<tr>
<td>263</td>
<td>1937</td>
<td>43°06’22.0&quot;</td>
<td>76°57’11.0&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>264</td>
<td>1923</td>
<td>43°05’50.1&quot;</td>
<td>76°57’30.0&quot;</td>
<td>birch</td>
</tr>
<tr>
<td>265</td>
<td>1923</td>
<td>43°05’53.0&quot;</td>
<td>76°57’23.1&quot;</td>
<td>apple</td>
</tr>
<tr>
<td>289</td>
<td>2509</td>
<td>43°03’44.6&quot;</td>
<td>76°59’19.3&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>293</td>
<td>2480</td>
<td>43°03’50.6&quot;</td>
<td>76°59’05.1&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>299 a</td>
<td>2312</td>
<td>43°04’19.8&quot;</td>
<td>76°59’11.1&quot;</td>
<td>willow</td>
</tr>
<tr>
<td>299 b</td>
<td>2312</td>
<td>43°04’19.8&quot;</td>
<td>76°59’11.1&quot;</td>
<td>aspen</td>
</tr>
<tr>
<td>301</td>
<td>2377</td>
<td>43°04’08.5&quot;</td>
<td>76°59’27.2&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>302</td>
<td>2395</td>
<td>43°04’06.4&quot;</td>
<td>76°59’26.9&quot;</td>
<td>rowan</td>
</tr>
<tr>
<td>304</td>
<td>1831</td>
<td>43°05’43.4&quot;</td>
<td>76°56’44.9&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>305</td>
<td>1864</td>
<td>43°05’35.0&quot;</td>
<td>76°56’52.9&quot;</td>
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<tr>
<td>309 a</td>
<td>1477</td>
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<td>76°54’53.1&quot;</td>
<td>hawthorn</td>
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<tr>
<td>310 a</td>
<td>1504</td>
<td>43°06’36.2&quot;</td>
<td>76°54’49.1&quot;</td>
<td>oak</td>
</tr>
<tr>
<td>310 b</td>
<td>1504</td>
<td>43°06’36.2&quot;</td>
<td>76°54’49.1&quot;</td>
<td>apricot</td>
</tr>
<tr>
<td>311</td>
<td>1556</td>
<td>43°06’31.4&quot;</td>
<td>76°54’43.8&quot;</td>
<td>birch</td>
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<tr>
<td>312</td>
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<td>43°06’29.5&quot;</td>
<td>76°54’43.8&quot;</td>
<td>spruce</td>
</tr>
<tr>
<td>313</td>
<td>1628</td>
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<td>76°54’46.6&quot;</td>
<td>apple</td>
</tr>
<tr>
<td>314</td>
<td>1667</td>
<td>43°06’21.9&quot;</td>
<td>76°54’47.9&quot;</td>
<td>pine</td>
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<tr>
<td>315 a</td>
<td>1703</td>
<td>43°06’18.3&quot;</td>
<td>76°54’49.6&quot;</td>
<td>rowan</td>
</tr>
<tr>
<td>315 b</td>
<td>1703</td>
<td>43°06’18.3&quot;</td>
<td>76°54’49.6&quot;</td>
<td>apple</td>
</tr>
<tr>
<td>315 c</td>
<td>1703</td>
<td>43°06’18.3&quot;</td>
<td>76°54’49.6&quot;</td>
<td>pine</td>
</tr>
</tbody>
</table>

**Isolation of fungi and preparation of medium**

Soil fungi were isolated by the serial dilution method (Polyksenova et al. 2004). Suspensions with standard level of dilution (1:10,000) were inoculated on agar. After the appearance of sporulation or signs of development of the mycelium, the isolates were transferred to a nutrient medium (potato sucrose agar). Petri dishes were labeled, sealed with paraffilm in order to avoid contamination, and incubated at 25°C.

Potato sucrose agar was made from 1800 g of potatoes per 4500 ml of water, 40 g of sucrose, 40 g of agar. The medium was autoclaved at 120°C for 30 minutes. A pinch of streptomycin (30 mg/ml) was added to the autoclaved medium to avoid the bacterial growth (Polyksenova et al. 2004).

**Growth of colonies**

Radial growth of fungal colonies on solid media was measured and calculated based on the average of perpendicular diameter measurements. Fungal growth patterns and their sizes were registered by a digital camera Canon 600E.

**Macroscopic and microscopic characterization of colonies and identification**

Fungal cultures were examined when they were 5–10 days old. Appearance on medium, color, texture, and growth rate of fungal colonies were analyzed.
For light microscopy, small pieces of fungal colonies (growing up to 10 days on potato sucrose agar) were cut, placed in a drop of distilled water on a microscope slide without any staining, examined and photographed using a photomicroscope Polyvar with Nomarski interference contrast optics. The diamentions of sporangiophores, sporangia, columellae, dimensions of sporangiospores and other fungal structures were recorded. Mean sizes were calculated and the differences between the isolates were tested. The obtained data were compared with the available literature.

Specimens were identified based on the taxonomic keys on soil Zygomycetes (Pidoplichko & Mil’ko 1971, Watanabe 2002). The systematics of the taxa were in accordance with Kirk et al. (2008) and they were listed in alphabetical order. Names of fungi are given according to Index Fungorum.

Occurrence frequency of fungal species was established according to the formula: \( A = \frac{B}{C} \times 100\% \), where \( A \) – occurrence frequency of fungal species; \( B \) – number of samples, in which the species has been detected; \( C \) – total number of investigated samples (Mirchink 1988).

Results

The Zygomycetes detected in the rhizosphere of various woody plants were identified as members of 9 species which fall into the genera *Cunninghamella, Absidia, Actinomucor, Mucor, Rhizopus, Piptocephalis*.

**Zygomycota** Moreau

**Mucoromycotina** Benny

**Mucorales** Dumort.

**Cunninghamellaceae** Naumov ex R.K. Benj.

*Zygomycota* Moreau

**Cunninghamellaceae** Naumov ex R.K. Benj.

**Mucorales** Dumort.

**Mucoraceae** Dumort.

**Cunninghamella** (Thaxt.) Thaxt. ex Blakeslee

In the hawthorn rhizosphere

- Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1477 m a.s.l., 43°06ʹ37.0ʺN, 76°54ʹ53.1ʺE, 29.08.2018, YV Rakhimova.

**Absidia** Lendn.

**Absidia spinosa** Lendn.

In the apple rhizosphere

- Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1628 m a.s.l., 43°06ʹ26.0ʺN, 76°54ʹ46.6ʺE, 29.08.2018, YV Rakhimova.

In the apricot rhizosphere

- Big Almaty gorge, North-West exposure slope, mixed forest, 1926 m a.s.l., 43°06ʹ27.1ʺN, 76°56ʹ15.9ʺE, 27.04.2018 YV Rakhimova; in the same gorge, slope of the Big Almaty peak, mixed forest, 1504 m a.s.l., 43°06ʹ36.2ʺN, 76°54ʹ49.1ʺE, 29.08.2018, AM Assylbek.

In the hawthorn rhizosphere

- Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1777 m a.s.l., 43°06ʹ37.0ʺN, 76°54ʹ53.1ʺE, 29.08.2018, AM Assylbek.

In the pine rhizosphere

- Big Almaty gorge, slope of the Big Almaty peak, the edge of the pine forest, 1667 m a.s.l., 43°06ʹ21.9ʺN, 76°54ʹ47.9ʺE, 30.08.2018, YV Rakhimova.

In the poplar rhizosphere

- Big Almaty gorge, North-West exposure slope, mixed forest, 1930 m a.s.l., 43°06ʹ28.7ʺN, 76°56ʹ21.2ʺE, 27.04.2018, LA Kyzmetova.

In the rowan rhizosphere

- Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1703 m a.s.l., 43°06ʹ18.3ʺN, 76°54ʹ49.6ʺE, 30.08.2018, YV Rakhimova.
In the spruce rhizosphere
Big Almaty gorge, North exposure slope, coniferous forest, 2377 m a.s.l., 43°04'08.5"N, 76°59'27.2"E, 09.08.2018, AM Assylbek; Ayusai gorge, South-Eastern exposure slope, coniferous forest, 1831 m a.s.l., 43°05'43.4"N, 76°56'44.9"E, 27.08.2018, LA Kyzmetova.

In the willow rhizosphere
Big Almaty gorge, North-West exposure slope, mixed forest, 2312 m a.s.l., 43°04'19.8"N, 76°59'11.1"E, 06.08.2018, UK Jetigenova.

**Actinomucor elegans** (Eidam) C.R. Benj. & Hesselt.
In the hawthorn rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1477 m a.s.l., 43°06'37.0"N, 76°54'53.1"E, 29.08.2018, AM Assylbek.

**Mucor fuscus** Bainier (*M. petrinsularis* Naumov)
In the rowan rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1703 m a.s.l., 43°06'18.3"N, 76°54'49.6"E, 30.08.2018, YV Rakhimova.

**Mucor mucedo** Fresen.
In the apple rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, coniferous forest, 1628 m a.s.l., 43°06'26.0"N, 76°54'46.6"E, 29.08.2018, YV Rakhimova; in the same gorge, mixed forest, 1703 m a.s.l., 43°06'18.3"N, 76°54'49.6"E, 30.08.2018, YV Rakhimova.

In the apricot rhizosphere
Big Almaty gorge, North-West exposure slope, mixed forest, 1926 m a.s.l., 43°06'27.1"N, 76°56'15.9"E, 27.04.2018 YV Rakhimova; in the same gorge, slope of the Big Almaty peak, mixed forest, 1504 m a.s.l., 43°06'36.2"N, 76°54'49.1"E, 29.08.2018, AM Assylbek.

In the aspen rhizosphere
Big Almaty gorge, North-West exposure slope, mixed forest, 2312 m a.s.l., 43°04'19.8"N, 76°59'11.1"E, 06.08.2018, UK Jetigenova.

In the birch rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, birch forest, 1556 m a.s.l., N43°06'31.4", E76°54'43.8", 29.08.2018, YV Rakhimova.

In the oak rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1504 m a.s.l., 43°06'36.2"N, 76°54'49.1"E, 29.08.2018, UK Jetigenova.

In the pine rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, the edge of the pine forest, 1703 m a.s.l., 43°06'18.3"N, 76°54'49.6"E, 30.08.2018, YV Rakhimova.

In the poplar rhizosphere
Big Almaty gorge, North-West exposure slope, mixed forest, 1930 m a.s.l., 43°06'28.7"N, 76°56'21.2"E, 27.04.2018, LA Kyzmetova; in the same gorge, mixed forest, 1957 m a.s.l., 43°06'23.8"N, 76°56'46.4"E, 27.04.2018, LA Kyzmetova.

In the spruce rhizosphere
Big Almaty gorge, South West exposure slope, coniferous forest, 2480 m a.s.l., 43°03'50.6"N, 76°59'05.1"E, 03.08.2018, YV Rakhimova; Ayusai gorge, South-Eastern exposure slope, coniferous forest, 1864 m a.s.l., 43°05'35.0"N, 76°56'52.9"E, 27.08.2018, LA Kyzmetova; Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1582 m a.s.l., 43°06'29.5"N, 76°54'43.8"E, 29.08.2018, AM Assylbek.

In the rowan rhizosphere
Big Almaty gorge, North exposure slope, coniferous forest, 2395 m a.s.l., 43°04'06.4"N, 76°59'26.9"E, 10.09.2018, YV Rakhimova.
In the willow rhizosphere
Big Almaty gorge, North-West exposure slope, mixed forest, 2312 m a.s.l., 43°04′19.8″N, 76°59′11.1″E, 06.08.2018, UK Jetigenova.

*Mucor silvaticus* Hagem (*M. hiemalis* f. *silvaticus* (Hagem) Schipper)
In the apple rhizosphere
Big Almaty gorge, mixed forest, 1923 m a.s.l., 43°05′53.0″N, 76°57′23.1″E, 27.04.2018, YV Rakhimova.

*Mucor strictus* Hagem
In the birch rhizosphere
Big Almaty gorge, birch forest, 1923 m a.s.l., 43°05′53.0″N, 76°57′23.1″E, 27.04.2018, YV Rakhimova.

*Rhizopus stolonifer* (Ehrenb.) Vuill. (*Rh. nigricans* Ehrenb.)
In the spruce rhizosphere
Big Almaty gorge, coniferous forest, 1937 m a.s.l., 43°06′22.0″N, 76°57′11.0″E, 27.04.2018, AM Assylbek.

Zoopagomycotina Benny
Zoopagales Bessey ex R.K. Benj.
*Piptocephalidaceae* J. Schröt.

*Piptocephalis arrhiza* Tiegh. & G. Le Monn.
In the apricot rhizosphere
Big Almaty gorge, slope of the Big Almaty peak, mixed forest, 1504 m a.s.l., 43°06′36.2″N, 76°54′49.1″E, 29.08.2018, AM Assylbek.

*Mucor mucedo* was observed as the most prevalent species in rhizosphere of apple, apricot, aspen, birch, oak, pine, poplar, spruce, rowan and willow. The colonies of isolates are white, dirty-white, beige, or gray, fluffy, very fast-growing (Table 2). Hyphae broad, nonseptate or sparsely septate. Sporangiophores erect; sporangia spherical and multispored, columellae of various shapes: cylindrical, rarely ovoid or pear-shaped, smooth (Fig. 2), (30.0–100.0 × 20.0–90.0) μm. The largest columellae are observed in fungi isolated from the rhizosphere of poplar and spruce (Table 3). Sporangiospores hyaline, cylindrical (Fig. 3), rounded at the ends, (10.0–14.0 × 6.5–8.0) μm. Rhizoid and zygospores were absent.

### Table 2 Macroscopic characters of different isolated Zygomycetes

<table>
<thead>
<tr>
<th>Fungal species</th>
<th>Color</th>
<th>Characters of colony</th>
<th>Growth rate</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Absidia spinosa</em></td>
<td>Grayish</td>
<td>Fluffy</td>
<td>Slightly yellowish</td>
</tr>
<tr>
<td><em>Actinomucor elegans</em></td>
<td>Unpainted, then gray</td>
<td>Fluffy</td>
<td>Slightly beige</td>
</tr>
<tr>
<td><em>Cunninghamella echinulata</em></td>
<td>Unpainted, later smoky</td>
<td>Arachnoid, fluffy</td>
<td>Beige</td>
</tr>
<tr>
<td><em>Mucor mucedo</em></td>
<td>Dirty white, beige or grey</td>
<td>Fluffy</td>
<td>Slightly greenish-yellowish</td>
</tr>
<tr>
<td><em>Mucor fuscus</em></td>
<td>Ash-colored</td>
<td>Fluffy</td>
<td>Unpainted or slightly beige</td>
</tr>
<tr>
<td><em>Mucor silvaticus</em></td>
<td>Pale ash</td>
<td>Fluffy</td>
<td>Slightly beige</td>
</tr>
<tr>
<td><em>Mucor strictus</em></td>
<td>Gray</td>
<td>Fluffy</td>
<td>Slightly yellowish</td>
</tr>
<tr>
<td><em>Piptocephalis arrhiza</em></td>
<td>Unpainted</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><em>Rhizopus stolonifer</em></td>
<td>Brownish-gray</td>
<td>Felt</td>
<td>Brownish</td>
</tr>
</tbody>
</table>
Table 3 Dimensions of columellae of different isolates of *Mucor mucedo*, µm

<table>
<thead>
<tr>
<th>Samples</th>
<th>Length of columella</th>
<th>Width of columella</th>
</tr>
</thead>
<tbody>
<tr>
<td>257</td>
<td>50.0–100.0</td>
<td>40.0–90.0</td>
</tr>
<tr>
<td>258</td>
<td>80.0–90.0</td>
<td>70.0–80.0</td>
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<td>289</td>
<td>50.0–100.0</td>
<td>30.0–90.0</td>
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<td>302</td>
<td>60.0</td>
<td>30.0–40.0</td>
</tr>
<tr>
<td>313</td>
<td>30.0–80.0</td>
<td>20.0–50.0</td>
</tr>
</tbody>
</table>

In addition to *Mucor mucedo* three species of this genus were noted. *Mucor fuscus* was isolated from the rowan rhizosphere. Colonies are ash-colored, fast-growing (Table 2). Sporangiophores hyaline, erect, branched, with short lateral branches bearing grayish brown spherical sporangia (Fig. 4). Columella cylindrical or ovoid, smooth. After the dehiscence of sporangia, hyaline, spherical (sometimes angular) sporangiospores release. Diameter of sporangiospores 8.0–10.5 µm.

*Mucor silvaticus* was found in the apple rhizosphere. Colonies are fast-growing (Table 2), fluffy, pale ash. Sporangiophores hyaline, erect, long; sporangia spherical. Sporangia first colorless, then brownish gray, spherical. Columella pale brown or colorless, spherical or ellipsoid. Sporangiospores hyaline, cylindrical. Dimensions of sporangiospores (5.5–6.5 × 2.5–4.0) µm.

*Mucor strictus*, isolated from the birch rhizosphere, was characterized by fast-growing, fluffy, gray colonies (Table 2). Sporangiophores hyaline, erect, long, sometimes curved or branched. Sporangia spherical or slightly flattened; columella pale brown, ovoid (Fig. 5). Diameter of spherical sporangiospores 5.0 µm.

*Absidia spinosa* was isolated from the rhizosphere of apple, apricot, hawthorn, pine, poplar, rowan, spruce and willow, and was observed less frequently than *Mucor*. Most often, the species is noted in the rhizosphere of spruce and apricot. Colonies are fast-growing, fluffy beige, grayish (Table 2). Rhizoids branched. Straight, brownish sporangiophores are situated in whorls on stolons (Figs 6, 7). Diameter of sporangiophores varies from 7.0 to 10.5 µm, while the diameter of sporangia – from 24.0 to 40.0 µm (Table 4). Dehiscence of sporangium takes place after maturation of globose sporangiospores. Columella hemispherical with a thin appendage (Fig. 8). Gametangia of the same size (Fig. 9). Zygospore, which develops after conjugation of gametangia, has simple, sickle-shaped curved appendages (Fig. 10).

Table 4 Dimensions of different isolates of *Absidia spinosa*, µm (a dash in a column indicates no measurements)

<table>
<thead>
<tr>
<th>Samples</th>
<th>Diameter of</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>sporangiophores</td>
</tr>
<tr>
<td>256</td>
<td>7.0–10.5</td>
</tr>
<tr>
<td>301</td>
<td>7.5</td>
</tr>
<tr>
<td>304</td>
<td>–</td>
</tr>
<tr>
<td>309 a</td>
<td>–</td>
</tr>
<tr>
<td>313</td>
<td>7.0–8.5</td>
</tr>
<tr>
<td>314</td>
<td>8.5–9.5</td>
</tr>
<tr>
<td>315 b</td>
<td>7.5–8.5</td>
</tr>
</tbody>
</table>

Only one species of *Cunninghamella (C. echinulata)* from hawthorn rhizosphere was identified. Colonies are fluffy, unpainted, white at first, later smoky (Table 2). Conidiophores straight, with verticillate or solitary branches. Conidia yellowish brown, spherical (Fig. 11), 10–15 µm in diameter, echinulate, with oil globules. Vesicles (Fig. 12) hyaline, pale brown, globose, subglobose or back pear-shaped, of different sizes. Apical vesicles (25.5–35.5 × 20.5–30.5) µm; vesicles on branches – (15.5–20.5 × 10.5–15.5) µm.
Figs 2–10 – Species of the genera *Mucor* and *Absidia*. 2 columella of *Mucor mucedo*. Scale Bar: 50 µm. 3 sporangiospores of *Mucor mucedo*. Scale Bar: 30 µm. 4 sporangium of *Mucor fuscus*. Scale Bar: 35 µm. 5 columella of *Mucor strictus*. Scale Bar: 60 µm. 6 sporangiophores on stolons *Absidia spinosa*. Scale Bar: 35 µm. 7 sporangium of *Absidia spinosa*. Scale Bar: 25 µm. 8 columella of *Absidia spinosa*. Scale Bar: 50 µm. 9 conjugation of gametangia of *Absidia spinosa*. Scale Bar: 25 µm. 10 zygospore of *Absidia spinosa*. Scale Bar: 40 µm.

*Actinomucor elegans* was isolated from the hawthorn rhizosphere. Colonies are fast-growing, fluffy, first colorless, then grayish. Hyphae colorless, septate, with few rhizoids. Sporangiophores pale brown, straight, sometimes with granular content, intensely branched. Sporangia spherical,
varying sizes: from 40-60 to 60-100 μm in diameter. After dehiscence of sporangium the columella seems colorless, slightly conical (Fig. 13).

*Piptocephalis arrhiza* was isolated from the apricot rhizosphere, together with species of the genus *Mucor*, it is their parasite. Hyphae uncolored, very thin, with rare septa; stolons absent. Merosporangiohores (Fig. 14) brownish, thin, repeatedly dichotomously branched; merosporangia simple, with 2–4 spores. Merosporangiospores pale brownish, smooth, cylindrical, (3.5–5.0 × 2.5–3.0) μm. Zygospore (Fig. 15) yellow, then rusty-brown, spherical with the star-shaped protrusions, 30.0–40.0 μm in diameter.

*Rhizopus stolonifer* was isolated from the spruce rhizosphere. Colonies are fast-growing, fluffy, brownish-gray. Rhizoids and stolons brown, transparent, well expressed (Fig. 16). Sporangiohores brown, erect, simple or branched, curved, bearing sporangia terminally. Sporangia dark brown to black, spherical, apparently subglobose after maturity, 100–200 μm in diameter; columella brownish to brown, globose (Fig. 17), 90–125 μm in diameter. Sporangiospores pale brown, spherical or ellipsoidal, (5.0–10.5 × 5.9–8.5) μm. Zygospores were not observed.

![Figs 11–17 – Species of the genera Cunninghamella, Actinomucor, Piptocephalis and Rhizopus.](image)


Eight species of Zygomycetes are found in the rhizosphere of mixed forest trees, three species are found in the soil of spruce forests (Table 5). Two species are common to mixed and spruce forests.
Table 5 Prevalence of fungal species (%) in different habitats (- – the species is absent, + – accidental species, occurrence frequency – A<30%, ++ – typical species, A>30%, +++ – dominant species, A>50%)

<table>
<thead>
<tr>
<th>Fungal species</th>
<th>Mixed and small-leaved forest</th>
<th>Coniferous forest</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1400–1700 m a.s.l.</td>
<td>1700–2000 m a.s.l.</td>
</tr>
<tr>
<td>Absidia spinosa</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td>Actinomucor elegans</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Cunninghamella echinulata</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Mucor mucedo</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td>Mucor fuscus</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Mucor silvaticus</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Mucor strictus</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Piptocephalis arrhiza</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Rhizopus stolonifer</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

The greatest number of soil fungi species is typical for mixed and small-leaved forest at the altitude range 1400–2000 m above sea level, much less of them in higher ranges (Table 5). Absidia spinosa and Mucor mucedo are dominant for these altitudes despite the fact that they are observed at all studied altitudes, except for coniferous forest for the interval 1900-2200 m above sea level.

The number of soil Zygomycetes varies in the rhizosphere of different forest species. The greatest number of soil fungal species (3 species) is observed in the rhizosphere of apple, apricot, birch, hawthorn, rowan and spruce (Fig. 18).

![Graph showing distribution of soil Zygomycetes in the rhizosphere of different forest species](image)

**Fig. 18** – Distribution of soil Zygomycetes in the rhizosphere of different forest species

**Discussion**

*Mucor* species are observed as the most prevalent species in rhizosphere of woody plants of Ile-Alatau Natural Park. Intensive development of *Mucor* fungi is associated with a high content of
poorly decomposed organic substances in the soil (Dyakov 2006, Berseneva et al. 2008). So, if in the rich forest soils to 25-40% of propagules of the considered fungi are *Mucor* fungi, then in the soils of arable land their number does not exceed 12-15% (Classen 2003). Some species are predominantly found in coniferous forest soils of Minsk upland, (Belomesyatseva & Shabashova 2004), in the rhizosphere of forest ecosystems of the European part of Russia (Kurakov & Semenova 2016), in the rhizosphere of various plants of the city of Lahore (Samina et al. 2017), in the rhizosphere of larch-spruce forests in the central part of the Dzhagdy ridge (Shumilova 2013), in the soils of Ukraine (Minter & Dudka 1996) and Poland (Budziszewska et al. 2010, Ruszkiewicz-Michalska et al. 2012, 2016). While *Mucor racemosus*, *M. nodosus*, and *M. mucedo* are found mainly in agricultural soils (Mirchink 1988).

*Absidia spinosa*, isolated from the rhizosphere of apple, apricot, hawthorn, pine, poplar, rowan, spruce and willow, is the next prevalent fungus in the studied rhizosphere samples from Ile-Alatau Natural Park. Fungi of the genus *Absidia* behave like fungi of the genus *Mucor* (Dyakov 2006, Berseneva et al. 2008). This may explain the high occurrence of *Absidia spinosa* in the rhizosphere of spruce in the forests of Ile-Alatau Natural Park. Species of the genus *Absidia* are also characteristic of the soil of coniferous forests of the Russian Far East (Egorova 2009, 2011, Egorova & Kovaleva 2012, Egorova et al. 2013).

Genus *Cunninghamella* is usually found in the soils of subtropical and tropical zones. This fungus is not found or poorly represented in the soils of the North (Egorova 2009, 2011, Egorova & Kovaleva 2012, Egorova et al. 2013, Shumilova 2013, Kurakov & Semenova 2016, Samina et al. 2017). In Kazakhstan, the genus *Cunninghamella* is represented by only one species *Cunninghamella echinulata* found once in the hawthorn rhizosphere in the forests of Ile-Alatau Natural Park. The same species is found in India (as *C. echinata*) (Koilraj et al. 1999). Another species *Cunninghamella elegans* is discovered in soils of Ukraine (Minter & Dudka 1996) and in dune, sand grassland of the Biebrza National Park, Poland (Ruszkiewicz-Michalska et al. 2012).

*Actinomucor elegans* isolated from the hawthorn rhizosphere in the forests of Ile-Alatau Natural Park is also found in the soils of Ukraine (Minter & Dudka 1996) and European part of Russia (Kurakov & Semenova 2016).

The genus *Rhizopus* is considered as a thermophilic and is more often found in the soils of the Southern zone (Dyakov 2006). However, *Rhizopus stolonifer* is common for the rhizosphere of forest ecosystems of Ile-Alatau Natural Park as well as Ukraine (Minter & Dudka 1996), Poland (Ruszkiewicz-Michalska et al. 2012, 2016), Russian Far East (Egorova 2009, 2011, Egorova & Kovaleva 2012, Egorova et al. 2013, Shumilova 2013), European part of Russia (Kurakov & Semenova 2016), Lahore city, Pakistan (Samina et al. 2017), and Arunachal Himalaya (Sharma et al. 2015), India, (Koilraj et al. 1999).

Mycophilic fungus *Piptocephalis arrhiza* is isolated from the apricot rhizosphere in mixed forest of Ile-Alatau Natural Park. Species of the genus *Piptocephalis* are obligate parasites of the members of *Mucorales* and they are commonly found on forest litter or in soil. *Piptocephalis* spp. are characterized by the production of dichotomously branching sporophores with cylindrical uniseriate merosporangia (Benjamin 1959, Mukerji 1968, Kirk 1978, Ho 2003, 2004, 2006a, 2006b). Currently, 24 species are described (Index Fungorum 2018). Of the 24 species only one species *Piptocephalis arrhiza* is found in Kazakhstan, while 3 species are registered on the territory of Biebrza National Park (Poland): *Piptocephalis fimbriata*, *P. lepidula*, *Piptocephalis* sp. (Ruszkiewicz-Michalska et al. 2012, 2016).

Thus, the species composition of the soil Zygomycetes of the Ile-Alatau Natural Park is distinguished by its originality, due to the variety of environmental conditions.

**Acknowledgements**

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